



## Study of possible seasonal effects on blood parameters of a local fish 'Schizothorax niger' in Kashmir valley

Ashiq Hussain Bhat

Assistant Professor, Department of Zoology, GDC, Women Anantnag, Jammu and Kashmir, India

### Abstract

Present work was carried out in order to find out the possible seasonal effects on the blood parameters of *Schizothorax niger* caught from the world-famous Dal Lake (Kashmir, India). Effects of the independent variables e.g., sex, weight and length values were determined seasonally. Analysis revealed that highest number of leukocytes was found in spring and lowest number was found in winter. Hemoglobin and Heamatocrit values were highest during the months of summer and lowest during winter. It was also seen that Males were having greater values for hemoglobin and Heamatocrit than females, whereas total leukocyte count in females was higher than males. It was also found that there was a positive correlation between Length/weight and Hemoglobin and Heamatocrit values where as a negative correlation between length or weight and Total leukocyte counts.

**Keywords:** schizothorax, leukocytes, heamatocrit values, correlation

### Introduction

Due to great importance of blood in the diagnosis of diseases in medicine, hematological and serological investigations are now employed on an increasing scale for diagnostic purposes in fish pathology. Hematology concerns with the cells present in the blood and serology deals with the constituents in the fluid part of the blood, such as proteins, minerals, carbohydrates, pigments, enzymes, hormones, immunoglobulins etc. Hematological investigations on fish have confirmed that the variations in the blood give indications of health conditions (Soirio, *et al* 1975; Hickey, 1976). Marked variations in hematological data, caused by the environment would necessitate an exact establishment of normal values of various environmental conditions and in different seasons of the year. In fisheries it is important to diagnose the illness of a fish by evaluating hematological data, particularly blood parameters (Rimish and Adanova, 1973). To picturize the actual hematological data is quite different because of marked internal and external variations sampling tecqniqs, laboratory techniques, seasonal variations, habitat size, sex, population density, oxygen content, pH, water, ammonia, feeding etc (Wilhelm filho *et al*, 1992; Hardig and Hogleung, 1984). Research on blood of cyprinus carpio had shown that blood parameters were affected by seasonal variations (Denton and Yousuf, 1975) [12]. In addition to this age, sex, weight and length of fish also affect the blood parameters of fish (Ezzat *et al*, 1973; Arda, 1974).

### Materials and Methods

In the present study 120 male and female specimens of *S. niger* were collected from Dal Lake with the help of local fisherman. Sex, weight and length of the specimens was male and females, 200-350gms and 25-35 cms respectively. Many

techniques were utilized for the collection of blood e.g. Heart punctures, severing of caudal peduncle (Blaxhall and Daisley, 1973; Hatting 1975) [7]. Blow was given on the head of the fish and the needle was inserted at an angle of 45° in to the heart and the blood was collected in to the glass tubes already containing EDTA (Ethylene diamine tetra acetic acid). Blood analysis was carried out very quickly after sampling. Different leukocyte counting was performed by transporting blood samples diluted with turck's solution with a leukocyte pipette on to a neubar's counting chamber and examined for leukocytes (Blaxhall and Daisley, 1973) [7]. Leishman staining was used to determine the percent leukocyte (Goel *et al*, 1981). The hemoglobin was determined according to the method of Haldane. Heamatocrit values were determined by the microheamatocrit (Jewet *et al*, 1991). The blood without EDTA was transferred in to Heamatocrit pipettes and centrifuged at 10,000×g rpm for 10 minutes and the ratio of the blood components in plasma was determined. The slides were examined under a binocular microscope with 100 x objectives to see the percentage of neutrophils, eosinophils, lymphocytes and monocyte.

### Results

Water quality plays a vital role in determining any hematological parameter of the fish. Water quality criteria (Tem.) of Dal Lake between January 2005 and December 2006 was different in different seasons. Total leukocyte count, oesinophil, neutrophil and monocyte percentages were significantly ( $p < 0.05$ ) higher for both sexes in spring -summer seasons than those of other seasons. Lymphocyte ratio also increased ( $p < 0.05$ ) in autumn-winter season. The amount of hemoglobin and Heamatocrit value increased significantly ( $p < 0.05$ ) in spring-summer season.

**Table 1:** Seasonal variations of total leukocytes and its types in males of S. Niger

Season/Temp.	No. of fishes	Total leukocyte count(x10/mm <sup>3</sup> )	Neutrophil%	Eosinophil%	Lymphocyte%	Monocyte%
Spring/16°C	15	14.00±0.20	27.10±0.50	1.65±0.1	63.40±1.50	11.20±0.30
Summer/28°C	15	13.60±0.50	26.40±0.40	1.05±0.2	63.70±1.20	10.50±0.50
Autumn/20°C	15	10.40±0.50	23.40±0.20	0.90±0.5	66.50±0.50	7.50±0.20
Winter/5°C	15	10.50±0.50	23.35±0.70	0.85±0.3	66.20±0.30	7.60±0.70

**Table 2:** Seasonal variations of total leukocytes and its types in females of S. Niger

Season/Temp.	No. of fishes	Total leukocyte count(x10/mm <sup>3</sup> )	Neutrophils%	Eosinophils%	Lymphocyte%	Monocyte%
Spring/16°C	15	19.00±0.50	34.00±0.70	1.02±0.02	55.05±0.30	15.50±0.17
Summer/28°C	15	17.50±0.30	28.50±0.50	0.80±0.50	60.02±1.20	13.70±1.09
Autumn/20°C	15	12.70±0.70	24.50±0.30	0.50±0.30	68.00±0.30	9.05±0.50
Winter/5°C	15	11.50±0.20	22.30±0.70	0.38±0.50	72.05±0.10	9.50±1.51

**Table 3:** Male dependent seasonal variations in Hemoglobin and Heamatocrit values

Season/Temp.	No. of fishes	Hemoglobin Density(g/dl)	Heamatocrit values (%)
Spring/16°C	15	8.50± 0.05	34.80± 0.30
Summer/28°C	15	10.20± 0.20	39.65± 0.45
Autumn/20°C	15	8.30± 0.70	30.50 ±0.37
Winter/5°C	15	7.02 ± 0.30	26.30 ± 0.70

**Table 4:** Female dependent seasonal variations in Hemoglobin and Heamatocrit values

Season/Temp.	No. of fishes	Hemoglobin Density(g/dl)	Heamatocrit values (%)
Spring/16°C	15	6.80± 0.08	27 ± 0.50
Summer/28°C	15	8.15± 0.05	29± 0.20
Autumn/20°C	15	5.70 ± 0.09	21± 0.10
Winter/5°C	15	5.20 ± 0.08	19.80± 0.20

**Table 5:** Comparison of Blood parameters between male and female S. niger Between March 2005 to Dec. 2006.

Blood parameters	Males	Females
Total leukocyte count	12.52± 1.50	15.50 ± 0.50
Leukocyte types (%)		
Neutrophils	24.20± 0.50	28.50± 0.30
Eosinophils	0.80 ± 0.01	0.50 ± 0.01
Lymphocytes	70.50± 0.80	65.20± 1.50
Monocyte	9.50 ± 1.00	11.50± 0.80
Hemoglobin Density(g/dl)	8.20 ± 0.50	7.02 ± 0.50
Heamatocrit values (%)	30.50± 0.70	24.50± 0.23

**Table 6:** Length-weight group dependent frequencies of hematological parameters of S. niger

Parameter				
No. of fishes	15	15	15	15
Length(cm)	17-23	24-30	31-35	35-39
Weight(gm)	55-150	150-280	282-350	350-380
Hemoglobin Density(g/dl)	6.02 ± 0.12	6.38± 1.50	7.02 ± 0.32	8.50 ± 0.3
Heamatocrit values (%)	19.50± 0.50	21.32± 1.62	23.60± 1.32	26.80± 1.35
TLC(x10/mm <sup>3</sup> )	25.52± 1.30	24.60± 0.80	22.80± 1.30	20.80± 0.50
Neutrophils (%)	11.60± 1.50	13.50± 0.20	14.60± 1.30	15.30± 0.20
Eosinophils (%)	0.50 ± 1.31	0.59 ± 0.05	0.70 ± 0.30	0.50 ± 0.70
Lymphocytes (%)	82.80± 0.50	80.50± 1.35	80.30± 1.35	75.30± 0.30
Monocyte (%)	10.50± 0.50	10.30± 0.50	11.20± 0.70	13.50± 0.40

The leukocyte count, neutrophils and monocyte percentages were compared regarding sex difference, they were found to be higher in females than males but the level of hemoglobin and Heamatocrit values and lymphocyte ratios were found to be higher in males than females. The amount of hemoglobin, Heamatocrit value, neutrophil and monocyte ratio also increased in length and weight but lymphocyte ratio decreased significantly.

**Discussion**

Although less work has been done on the hematology of S.niger there are few reports of normal blood values and

published values are severely limited by low fish numbers. Annual changes in blood parameters showed a clear increase in spring-summer period and a decrease in autumn-winter period. The leukocyte numbers and Neutrophil ratios for female specimen were found to be higher compared to male specimen during the reproduction period (March-August). The same trend occurred with the values of hemoglobin, heamatocrit, neutrophil and monocyte percentages. Our results are in accordance with other results of (Denton and yousuf, 1975; Ezzat, A.A, *et al* 1974; Beelen, R. *et al*, 1998; Pitomberia, M.S *et al* 1970; Azizo-lu and Cengizler, 1996)<sup>[12]</sup>. When sex was taken into consideration, the amount of

hemoglobin, Hematocrit and lymphocyte values for male fish were significantly higher than for female fish. Monocyte ratio, neutrophil and leukocyte values were higher in females at reproduction periods than in males. So, our results are in accordance with the results of Ezzat *et al* 1974; Murray 1984 [20]; Terao *et al* 1984; Lumsden. J.H 1998. Length to weight independent variables were determined to cause an increase in hemoglobin, hematocrit, neutrophil and Monocyte values but the ratio of lymphocytes decreased to a moderate level. The length weight variables on blood parameters (lymphocyte formation) is an essential component of immune system in the early stages of growth period.

To conclude, the large variation in hematological parameters obtained emphasizes the need of more extensive study. The changes in micro-environment and the macro-environment have a direct effect on the blood components of fish which makes the comparison difficult to be made. Evaluation of hematological analytes will enhance the culture of fish by facilitating early detection of infectious diseases and identification of sub lethal conditions affecting production performance this will contribute to more specific, timely and effective disease treatment in the future. Therefore, we are of this opinion that there should be separate data collection and comparison from healthy and unhealthy fish to obtain reliable hematological data. These results of our study may be used as reference values of blood parameters for other fishes.

## References

1. Alcorn SW, Murray AL, Pascho RJ. Effects of rearing temperature on immune functions in sockeye salmon (*Oncorhynchus nerka*). *Fish & Shellfish Immunology*. 2002; 12:303-334.
2. Alexander N, *et al*. Hematological characteristics of albacore, *Thunnus alalunga* (Bonnaterre), and skipjack, *Katsuwonus pelamis*. *Jour. Fish Biol.* 2006; 16:383-395.
3. Alvarez-Pellitero, *et al*. Some blood parameters in sea bass, *Dicentrarchus labrax*, infected by bacteria, virus and parasites. *Jour. Fish Biol.* 2005; 31:259-261.
4. Bennetm MF, *et al*. Effects of cold shock on the distribution of leucocytes in goldfish, *Carassius auratus*. *J Comp. Physiol.* 2005; 98:213-216.
5. Boyar HC. Blood cell types and differential cell counts in Atlantic herring, *Clupea harengus harengus*. *Copeia*. 2005; 2:463-465.
6. Burrows F, *et al*. Blood leucocytes of the turbot, *Scophthalmus maximus* L. *Aquaculture*. 2006; 67:214-215.
7. Blaxhall PC, Daisley KW. Routine haematological methods for use with fish blood. *Journal of Fish Biology*. 1973; 5:571-604.
8. Bridges DW, Cech Jr J, Pedro DN. Seasonal hematological changes in winter flounder, *Pseudopleuronectes americanus*. *Transactions of the American Fisheries Society*. 1976; 105:596-600.
9. Burrows AS, Fletcher TC. Blood leucocytes of the turbot *Scophthalmus maximus* L. *Aquaculture*. 1987; 67(2):14-215.
10. Catton J. Blood cell formation in certain teleost fishes. *Blood*. 1951; 61:39-60.
11. Conroy DA. Studies on the haematology of Atlantic salmon (*Salmo salar* L.). *Symposia of the Zoological Society of London*. 1972; 30:101-127.
12. Denton JE, Yousef MK. Seasonal changes in haematology of rainbow trout, *Salmo gairdneri*. *Comparative Biochemistry and Physiology SIA*. 1975; 15:1-153.
13. Dunn SE, Murad A, Houston AH. Leucocytes and leucopoietic capacity in thermally acclimated goldfish, *Carassius auratus* L. *Journal of Fish Biology*. 1989; 34:223-230.
14. Ellis AE. Leucocytes and related cells in the plaice *Pleuronectes platessa*. *Journal of Fish Biology*. 1976; 8:143-156.
15. Enomato Y. On some notes about the fluctuations of the blood leucocyte numbers of the cultured fish. *Bull. Tokai reg. Fish. Research Lab.* 2006; 57:137-177.
16. Kekic H, Ivanc A. A new direct method for counting fish blood cells. *Ichtyo-logia*. 1982; 14:55-58.
17. Kori-Siakpere O. Haematological characteristics of *Clarias isteriensis* Sydenham. *Journal of Fish Biology*. 1985; 27:259-263.
18. Korzhuev PA, Alyorkrinskaya CO, Dgora SN. Characteristics of the blood in young and adult *Salmo salar* (Salmonidae). *Journal of Ichtyology*. 1982; 22:112-120.
19. Lehmann J, Stiirenberg FJ. Haematologisch serologisch Suhstratunter-suchungen an der Regenbogenforelle (*Salmo gairdneri* Richardson). *Krefeld-Miils: Kaltenmeier*. 1975, 901-911.
20. Murray SA. Hematological study of the bluegill, *Lepomis macrochirus* Raf. *Comparative Biochemistry and Physiology*. 1984; 78A:787-791.
21. Peters G, Schwarzer R. Changes in hemopoietic tissue of rainbow trout under influence of stress. *Diseases of Aquatic Organisms*. 1985; 1:1-10.
22. Pickering AD. Changes in blood cell composition of the brown trout *Salmo trutta* L., during the spawning season. *Journal of Fish Biology*. 1986; 29:335-347.
23. Preston A. Red blood values in the plaice (*Pleuronectes platessa* L.). *Journal of the Marine Biological Association of the United Kingdom*. 1960; 39:68 1-687.